

**Genetic and morphometric comparison of two isolated populations of *Barilius bendelisis* (Cypriniformes: Cyprinidae) from the Indian Himalayas**  
**Comparación genética y morfométrica de dos poblaciones aisladas de *Barilius bendelisis* (Cypriniformes: Cyprinidae) de los Himalayas indios**

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### Abstract

*Barilius bendelisis* (Hamilton) inhabits cold water drainages of the Himalayan region, occurring near the stream banks. It is an important component of the diet of rural population of Uttarakhand and Jammu. Despite the aquaculture importance of *B. bendelisis*, no extensive molecular characterization from two geographically isolated rivers, Alaknanda and Chenab, has been conducted. In order to study those aspects, 567 samples of *B. bendelisis* were analysed and collected from these tributaries of two geographically isolated rivers between March of 2015 and April 2017. The morphometric data were analysed by means of truss analysis using tpsDig2 and PAST, whereas the genetic characterization was performed using the COI gene. In truss analysis 14 landmarks resulting in 90 measurements were studied from the digitized images of the sampled specimens. In total 23 measurements exhibited significant differences among the populations of *B. bendelisis*. The principal component analysis (PCA) generated seven components explaining- 93.15 % of total variance. Discriminant function analysis (DFA) revealed that 83.8 % of the specimens were classified into their original populations. Truss based morphometry and Maximum likelihood type of phylogenetic tree revealed heterogeneity between the two geographically isolated populations of *B. bendelisis*.

**Key words:** *Barilius bendelisis*; truss analysis; PAST; Cypriniformes; morphometric characterization; molecular characterization.

### Resumen

*Barilius bendelisis* (Hamilton) habita las aguas frías de de las riberas de los ríos del Himalaya. Es un componente importante de la dieta de la población rural de Uttarakhand y Jammu. A pesar de la importancia de *B. bendelisis* en la acuicultura, no se reportan caracterizaciones moleculares de los ríos geográficamente aislados, Alaknanda y Chenab. Para estudiar estos aspectos, analizamos 567 muestras de *B. bendelisis* que se recolectaron de los afluentes de estos dos ríos entre marzo 2015 y abril 2017. Analizamos los datos morfométricos por medio del análisis truss utilizando tpsDig2 y PAST y para la caracterización genética utilizamos el gen COI. En el análisis truss 14 puntos de referencia que resultaron en 90 medidas se analizaron de imágenes digitalizadas de especímenes

muestreados. Un total de 23 medidas mostraron diferencias significativas entre las poblaciones de *B. bendelisis*. El análisis de componentes principales (PCA) generó siete componentes explicando el 93.15 % del total de la varianza. El análisis de función discriminante (DFA) reveló que el 83.8 % de los especímenes fue clasificado dentro de sus poblaciones originales. La morfometría basada en el análisis de truss y el árbol filogenético de máxima verosimilitud revelaron heterogeneidad entre las dos poblaciones geográficamente aisladas de *B. bendelisis*.

**Palabras clave:** *Barilius bendelisis*; análisis truss; PAST; Cipriniformes; caracterización morfométrica; caracterización molecular.

## Introduction

*Barilius bendelisis* (Hamilton, 1807) commonly called as “Indian hill trout” is a member of the family Cyprinidae that occurs in shallow and cold waters of rivers and streams of the Himalayan region, northeastern India and several parts of Uttarakhand. This fish species is considered a fishery resource and also is found in the lentic and lotic water bodies where the exotic carp (*Cyprinus carpio* and *Hypophthalmichthys molitrix*) and Indian major carp (*Labeo rohita*, *L. calbasu*, *L. bata*, *Cirrhinus miragla* and *C. reba*) cannot be raised successfully (Sahoo, Saikia, & Das, 2009).

*Barilius bendelisis* reaches a maximum length of 22.7 cm and is characterized by dark blue bars on the body, an elongated and compressed body and a dorsal fin inserted posterior to the mid length of the body (Rahman, 1989). Adults of *Barilius bendelisis* inhabit base hills streams and rivers with pebbles at bottom (Talwar & Jhingran, 1991; Menon, 1999). As per IUCN (2012) the species has been placed as least concerned but in the future anthropogenic activity, causing habitat destruction and over exploitation are the main threats to this species. The overuse of *B. bendelisis* in the rural fishing and aquarism is negatively impacting on its populations in the Himalayan region in the last years (Sah, Barat, Pande, Sati, & Goel, 2011). The species is mainly identified by means of morphological features, which sometimes are not useful to correctly identify all the existing populations because of its inherent high variability (Victor, Hanner, Shivji, Hyde, & Caldwell, 2009).

Interspecific variation shown by using COI barcodes in several studies is higher as compared to the intraspecific variation (Hubert et al., 2008; Rock, Costa, & Walker, 2008; Swartza, Mwalea, & Hanner, 2008; Steinke, Zemlak, & Hebert, 2009; Ward, Hanner, & Hebert, 2009). The present study aimed at to compare and illustrate morphometrics and molecular characters of *Barilius bendelisis* from the two geographically isolated tributaries of Alaknanda and Chenab Rivers.

In spite of high aquaculture scope, very limited morphometric and molecular studies have been conducted for this fish species. The Truss Network Analysis overcomes the disadvantages of manual morphometry and this method produces a more systematic geometric characterization of fish shape and describes inter-specific shape differences with high resolution (Humphries et al., 1981; Strauss & Bookstein, 1982). Molecular characterization supports morphometric analysis. Therefore, the present study was designed to characterize and compare the fish morphometrically and at molecular level from two isolated rivers of Western and Central Himalaya.

## Materials and methods

## Collecting and identification of samples

The Chenab River is the largest tributary of the Indus River of Western Himalaya while the Alaknanda River is the largest tributary of Ganga River of Central Himalaya. Fresh specimens of *B. bendelisis* were captured using various fishing gears (cast nets, gill nets and traditional Thali method) with the help of local fishermen from March 2015 to April 2017. A total of 267 specimens were collected from three different tributaries of the Chenab River Jhajjar, Dhudhar and Jhuni and 300 specimens were collected from other three tributaries of the Alaknanda River: Dugadda, Khankhra and Khandah Gad (Table 1). All the specimens were collected before the breeding season and after the spawning season. A solution of 10 % of formalin was utilized to fix and preserve the collected samples that were morphometrically analysed. Some samples were stored in 95 % ethanol to perform the DNA analysis. The captured specimens were identified with the help of taxonomic keys of Day (1878), Talwar and Jhingran (1991), Mirza (1991), and Kullander, Fang, Delling, and Ahlander (1999). The present specimens of *B. bendelisis* were deposited in the museum of the Department of Zoology and Biotechnology, HNB Garhwal University, Srinagar (Garhwal).

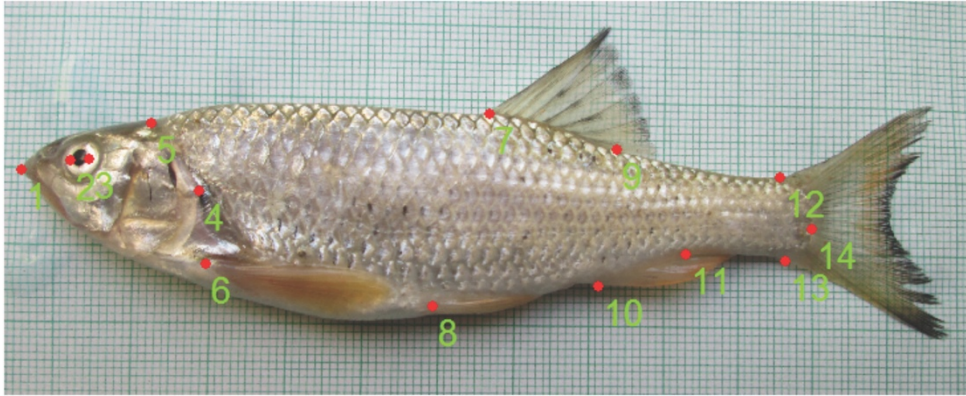
TABLE 1  
GPS coordinates and number of samples of *Barilius bendelisis* collected from the Indian Himalaya

Variables	Sampling sites					
	Alaknanda River (Central Himalaya)			Chenab River (Western Himalaya)		
	Dugadda	Khankhra	Khandah	Dhudhar	Jhajjar	Jhuni
Latitude °N	30°15'31"	30°14' 32"	30°11'47"	32°55' 15"	32°52' 25"	32°53' 51"
Longitude °E	78°43'7.89"	78°55' 36"	78°46' 34"	75°01' 42"	74°59' 54"	74°57'17"
Number of samples collected	98	94	108	91	89	87

## Morphological analysis

**Digitalization of specimens:** The specimens were firstly cleaned in running water, soaked by blotting paper and then placed on a flat platform with graph paper as background for digital images. To make the insertion points visible, the fins were erected and each individual was labelled with a specific code for own identification. Photographs were taken by means of a digital camera of (Nikon D3400). To avoid errors during the image capture, all photographs were taken by a single person from the same angle and height.

**Truss analysis:** A set of three software platforms, tpsUtil, tpsDig2 v2.1 (Rohlf, 2006) and Paleontological Statistics (PAST) (Hammer, Harper, & Ryan, 2001) was used to measure the truss distances of the digital images. The truss analysis was based on 14 landmarks (Fig. 1) (Strauss & Bookstein, 1982; Mir, Sarkar, Dwivedi, Gusain, & Jena, 2013).



**Fig. 1.** Schematic image of *B. bendelisis* showing the 14 landmarks used to compare among the populations. 1 tip of snout; 2 anterior border of eye; 3 posterior border of eye; 4 posterior border of operculum; 5 forehead (end of frontal bone); 6 pectoral fin origin; 7 dorsal fin origin; 8 pelvic fin origin; 9 dorsal fin termination; 10 origin of anal fin; 11 termination of anal fin; 12 dorsal side of caudal peduncle, 13 ventral side of caudal peduncle; 14 termination of lateral line.

**Statistical procedures:** Size dependent variation was conducted by using an allometric method suggested by Elliott, Haskard, and Koslow (1995) as “ $M_{adj} = M (L_s/L_0)^b$ ”, where  $M$  is original measurement,  $M_{adj}$  is the size adjusted measurement,  $L_0$  is the standard length of the fish,  $L_s$  the overall mean standard length, and  $b$  was estimated for each character from the observed data as the slope of the regression of  $\log M$  on  $\log L_0$  using all fish from every group. The significance of the correlation between transformed variables and standard length was tested using SPSS and the results derived from the allometric method were confirmed (Turan, 1999). Ninety morphometric characters were subjected to a univariate analysis of variance (ANOVA) to compare the populations of the different sampled sites of *B. bendelisis*. Then the significant variables ( $P < 0.05$ ) were subjected to Factor analysis. A maximum likelihood method was used to extract the factors. The cumulative variance explained by the factors and loading of significant traits were retained. The retained factors were subjected to varimax rotation procedure to identify the variables demonstrating high loading for a given factor as described by Hatcher (1994). The Wilks  $\lambda$  was used to compare the difference between all groups. Additionally, the significant measurements were also subjected to the discriminant function analysis (DFA) to calculate percentage of correctly classified fish. Statistical analysis for morphometric data were performed using the SPSS vers.20 and Microsoft Excel 2007.

## DNA analysis

**Isolation of DNA, amplification and sequencing:** Approximately 50 mg tissue was used for genomic DNA isolation following Phenol-Chloroform -Isoamyl alcohol method of Sambrook Fritsch, and Maniatis (2001). Genomic DNA quality was checked using 0.8 % agarose gel electrophoresis (Genei, Banaglore) while the qualitative and quantitative estimation was done by observing the bands in ultraviolet light on UV transilluminator (BioRad, USA) with UV shield and UV protective goggles. The genomic DNA was stored at  $-20\text{ }^{\circ}\text{C}$  for future use. The mitochondrial gene cytochrome c oxidase subunit I (COI) was amplified with primers FishF1-CGTATTTTTTGGCTTGAGCCG and FishR-AATAAGCCCAAACCCGGGAA in a 50 ml volume with 100 ng template DNA, 10 pmol of each specific primer, 250 mM of each

dNTPs, 1.0 U of Taq DNA polymerase and 1\_Taq buffer containing 1.5 mM MgCl<sub>2</sub>. The PCR conditions were consisted of initial denaturation at 95 °C for 3 minutes, 30 cycles, cycle denaturation at 95 °C for 20 seconds, cycle annealing at 47 °C for 20 seconds, cycle extension at 72 °C for 45 seconds, final extension at 72 °C for 10 minutes. To visualize the PCR products after that, a 1.2 % agarose gel was used, the amplicons were purified and sequenced in both directions commercially in automated genetic analyzer ABI 3730 (Applied Biosystem, Carlsbad, CA) following Bigdye terminator v.1.1 cycle Sequencing Kit (Applied Biosystem, Carlsbad, CA). Simplicity and un-ambiguity were observed among the sequences of the mitochondrial region. A total of 18 sequences of COI gene of *B. bendelisis* were analysed as follow: nine samples were obtained from the three collecting sites at the Chenab River and nine samples were obtained from the other three sites at the Alaknanda River. The sequences were submitted to the NCBI Genbank having accession number MF445031, MF445032, MF445033, MF445028, MF445029, MF445030, MF445034, MF445035, MF445036 for *B. bendelisis* of Chenab River and MG797659, MG797660, MG797661, MG797662, MG797663, MG797664, MG797665, MG797666, MG797667 for *B. bendelisis* of Alaknanda River.

**Sequence analysis:** the raw sequences were initially trimmed and aligned using the Clustal W (Thompson, Gibson, Plenwniak, Jeanmougin, & Higgins, 1997). The COI sequence confirmation was carried out using BLAST program of NCBI.

### Estimation of Genetic distance and Phylogenetic relationship

The number of polymorphic sites (including the number of constant, variables, and parsimony informative sites) and nucleotide composition were determined by DnaSpver 3 (Rozas & Rozas, 1999). The sequence divergence, determination of pairwise evolutionary distance among haplotype and Maximum likelihood tree was constructed by using MEGA 6.0 (Tamura, Stecher, Peterson, Filipski, & Kumar, 2013). Maximum likelihood tree with bootstrap value 100 was constructed on the basis of COI data and taking some other sequences of genus *Barilius* (*Barilius ngawa* KJ909413.1, *Barilius barna* JN965190.1, *Barilius gatensis* KX230845.1, *Barilius bakeri* KX230847.1, *Barilius barila* KY867678.1, *Barilius vagra* KC791629.1, *Barilius bendelisis* MF445035, *Barilius bendelisis* FJ459420 and *Schizothorax richardsonii* KR809777) from NCBI- GenBank just to increase the resolution among the species.

## Results

Ninety variables were obtained from the 14 landmarks using the truss analysis (Fig. 1). The ANOVA for 90 parameters extracted 23 significant characters ( $P < 0.05$ ). Principal component analysis of 23 truss measurements extracted 7 factors with eigen-value  $> 1$ , explaining 93.15 % of variance (Fig. 2). The first principal component (PC1) accounted for 37.11 % of variation followed by PC2 15.57 %, PC3 12.02 %, PC4 9.7 %, PC5 7.77 %, PC6 6.20 % and PC7 4.65 % are shown in Table 2 and their loadings are shown in Table 3.

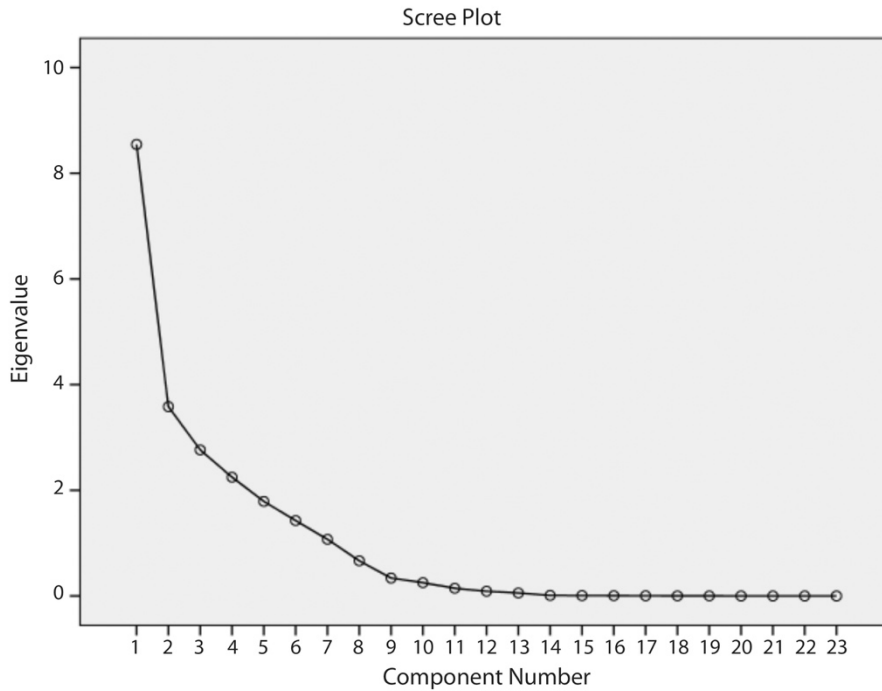
TABLE 2  
Total variance explained on the basis of PCA

Componen	Initial Eigen values	Extraction sums of squared loadings
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t	Total	% of variance	Cumulative %	Total	% of variance	Cumulative %
PCA1	8.546	37.1555	37.1555	8.546	37.115	37.115
PCA2	3.582	15.575	52.730	3.582	15.575	52.730
PCA3	2.765	12.020	64.750	2.765	12.020	64.750
PCA4	2.247	9.769	74.519	2.247	9.769	74.519
PCA5	1.789	7.777	82.296	1.789	7.777	82.296
PCA6	1.426	6.201	88.497	1.426	6.201	88.497
PCA7	1.070	4.654	93.151	1.070	4.654	93.151

TABLE 3  
Extracted principal components with significant loadings

	Component						
	PC 1	PC 2	PC 3	PC 4	PC 5	PC 6	PC 7
VAR 1-2	-0.796	.516	-.179	.037	.039	-.044	.103
VAR 1-3	-0.802	.528	-.106	.041	.021	-.063	.108
VAR 1-4	-0.798	.500	.102	.056	-.034	-.015	.084
VAR 1-5	-0.654	.518	.008	.108	-.050	.110	.197
VAR 1-6	-0.565	.131	.644	.202	-.241	.290	-.132
VAR 1-7	-0.232	.381	.339	-.293	-.086	-.385	.031
VAR 1-9	0.092	.259	.397	-.262	.802	.202	.023
VAR 1-10	0.011	.193	.456	.561	.120	-.583	-.256
VAR 1-11	0.325	.807	-.162	.386	-.030	.192	-.088
VAR 1-13	0.201	-.069	.260	.572	.068	-.058	.731
VAR 2-6	-0.192	.034	.697	.213	-.289	.502	-.132
VAR 2-7	0.110	.138	.413	-.257	.841	.132	-.011
VAR 2-9	0.578	.425	.408	-.444	-.286	-.060	.108
VAR 2-10	0.488	-.080	.475	.458	.085	-.475	-.253
VAR 2-11	0.792	.438	-.076	.258	-.031	.249	-.095
VAR 2-13	0.700	-.374	.242	.280	.007	.043	.464
VAR 2-14	0.814	-.476	.121	-.138	-.030	.109	-.044
VAR 3-9	0.618	.393	.377	-.443	-.290	-.055	.099
VAR 3-11	0.816	.394	-.094	.240	.006	.253	-.102
VAR 4-9	0.785	.264	.242	-.353	-.225	-.089	.055
VAR 4-11	0.871	.222	-.199	.250	.013	.162	-.120
VAR 6-9	0.654	.492	-.224	-.279	-.021	-.270	.132
VAR 6-11	0.652	.401	-.544	.205	.192	-.074	-.003



**Fig. 2.** Scree plot showing loading of significant variables.

In the DFA five discriminant functions (DFs) were obtained. The first discriminant function accounted for 48.3 %, second, third, fourth and fifth discriminant functions accounted for 24.3, 17.7, 6.5 and 3.1 % of the total variance respectively (Table 4). The DF1 vs DF2 plot explained variance among the samples and formed six groups. Group 1, 2, 3 from the Alaknanda River and group 4, 5, 6, from the Chenab River. The plot shows that these groups are clearly separated towards left and right showing their isolation. However, the group 4, 5, 6 from the Chenab River shows resemblance with the group 2 of the Alaknanda River (Fig. 3). The results obtained from the truss-based morphometrics indicated that the *B. bendelisis* have phenotypic heterogeneity between the two geographically isolated rivers of the Western and Central Himalayan regions. Individual’s classification into their original population varied between 79.1 and 92.6 % by discriminant function analysis being highest in the Khankhra gad drainage, a tributary of the Alaknanda River. In total 83.8 % of the examined individuals could be classified into their original groups (Table 5).

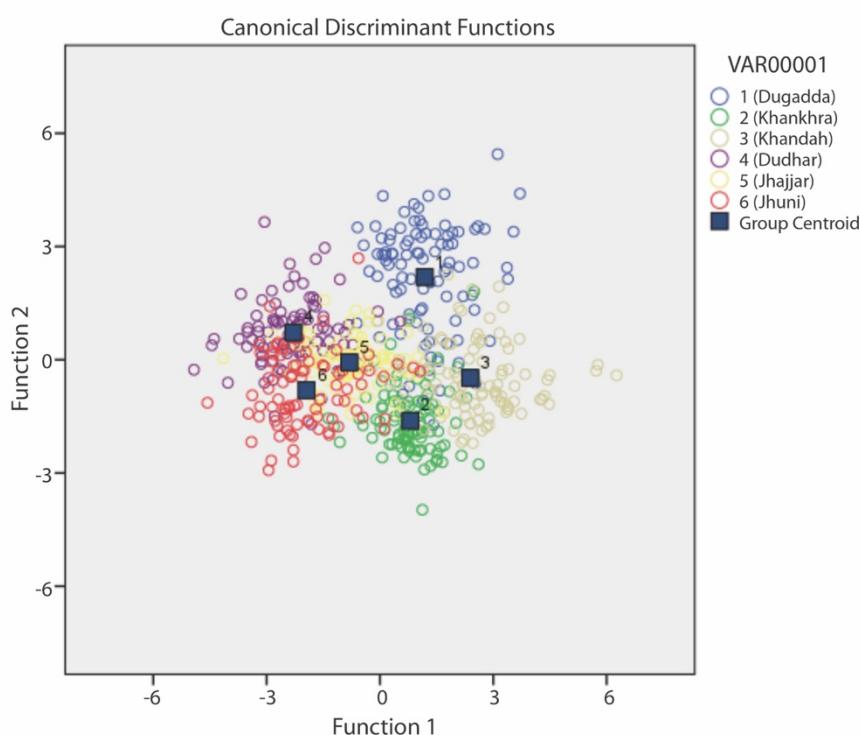
TABLE 4  
Total variance explained on the basis of DFA

Component	Eigen value	% variance	Cumulative %	Canonical correlation
DFA1	2.995 <sup>a</sup>	48.3	48.3	0.866
DFA2	1.507 <sup>a</sup>	24.3	72.6	0.775
DFA3	1.099 <sup>a</sup>	17.7	90.4	0.724
DFA4	.404 <sup>a</sup>	6.5	96.9	0.536
DFA5	.193 <sup>a</sup>	3.1	100	0.402

TABLE 5  
Percentage of specimens in each group and their percentage after cross validation for morphometric measurements of *Barilius bendelisis* from two rivers across Indian Himalaya

Sampling sites	Predicted group membership						
	Alaknanda River			Chenab River			Total
	Dugadda	Khankhra	Khandah	Dudhar	Jhajjar	Jhuni	
Dugadda	81	6	7	0	3	1	98
Khankhra	2	87	4	0	1	0	94
Khandah	8	4	90	0	6	0	108
Dudhar	0	0	0	72	11	8	91
Jhajjar	0	3	0	10	74	2	89
Jhuni	0	3	0	12	1	71	87
Cross validated results (%)							
	Dugadda	Khankhra	Khandah	Dudhar	Jhajjar	Jhuni	Total
Dugadda	82.7	6.1	7.1	0.0	3.1	1.0	100.0
Khankhra	2.1	92.6	4.3	0.0	1.1	0.0	100.0
Khandah	7.4	3.7	83.3	0.0	5.6	0.0	100.0
Dudhar	0.0	0.0	0.0	79.1	12.1	8.8	100.0
Jhajjar	0.0	3.4	0.0	11.2	83.1	2.2	100.0
Jhuni	0.0	3.4	0.0	13.8	1.1	81.6	100.0

83.8 % of original grouped cases correctly classified.



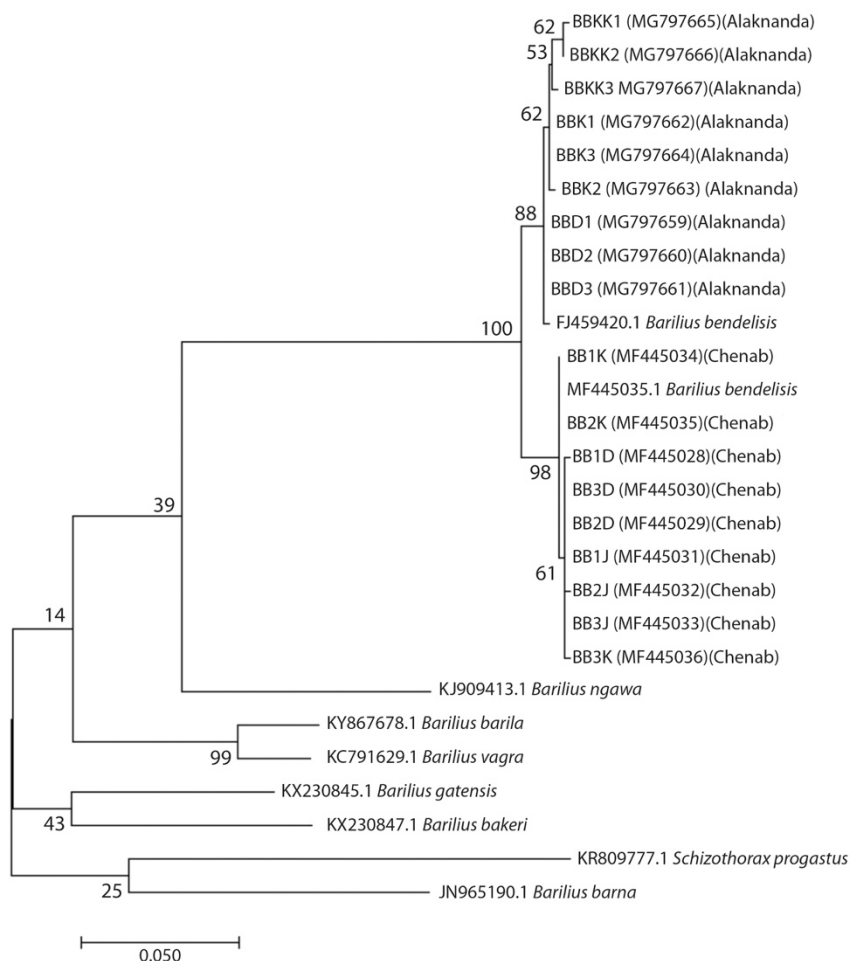
**Fig. 3.** Discriminant analysis plot of *Barilius bendelisis* from two isolated rivers of Indian Himalaya.

A cross-validation test using leave one out procedure also revealed that the samples were correctly classified into their original populations. A slight intermingling was observed among the samples of streams of the Western Himalayan River (Chenab) and Central Himalayan River (Alaknanda) (Fig. 3).



Sequence alignment resulted into 640-642 bp per sequence after exclusion of the primer sequence and equal length alignment. No insertions, deletions or stop codons were observed in any of the sequence and all the amplified sequences derive from a functional mitochondrial COI gene. The COI sequence analysis revealed average nucleotide composition in *B. bendelisis* from the Alaknanda River as 30.8 (T), 27.2 (C), 22.5 (A), and 19.6 (G) while 30.0 (T), 27.8 (C), 24.8 (A), and 19.9 (G) in case of *B. bendelisis* from the Chenab River.

COI gene revealed seven variable polymorphic sites and six parsimony informative sites in the specimens from the Alaknanda River, whereas four variable polymorphic sites and one parsimony informative site in the specimens from the Chenab River. A total of 11 different haplotypes were observed in the groups under study of *B. bendelisis*, six haplotypes were found among the populations from the Alaknanda River and 5 haplotypes for the population of the Chenab River, which indicate a gene flow within the tributaries of both rivers. The haplotype gene diversity Hd of the studied specimens from the Alaknanda River was higher (0.889) than that of the studied specimens from the Chenab River (0.806). A similar pattern was found for the value of nucleotide diversity (Alaknanda region = 0.00475) (Chenab region = 0.00175). In the Maximum likelihood tree, constructed on the basis of COI data and taking some other sequences of genus *Barilius* and *Schizothorax richardsonii* (Gray) from NCBI-GenBank just to increase the resolution among the species, formed two major clusters of *B. bendelisis* one from the Alaknanda River and other from the Chenab River (Fig. 4). The maximum value of genetic distance between the *B. bendelisis* of both the rivers was 0.031.



**Fig. 4.** Maximum likelihood tree showing phylogenetic relationship among *Barilius bendelisis* species and other cyprinids based on COI mitochondrial sequences.

## Discussion

Among vertebrates, fishes show higher degree of variation within and between populations than other vertebrates, and are more prone to environmentally induced morphological variation (Wimberger, 1992). Such variation in morphology is commonly due to isolation of populations. The results of present study also revealed that *B. bendelisis* have phenotypic heterogeneity between the two Himalayan Rivers (Alaknanda and Chenab) which are geographically isolated. Karakousis, Triantaphyllidis, and Economidis (1991) reported DFA to be useful method to distinguish different stocks of the some species. Mir, Saxena, Patiyal, and Sahoo (2015) analysed morphometric heterogeneity among different populations of *Barilius bendelisis* in four rivers from Central Indian Himalaya by applying DFA. They analysed three discriminant functions (DFs) which accounted for 98.9, 0.6 and 0.5 % of total variation for DF1, 2 & 3. of the total variation, the second discriminant function (DF II) and third discriminant function (DF III) accounted for 0.6 and 0.5 % respectively, among the populations. Whereas in the present study five discriminant functions were extracted and accounted for 48.3, 24.3, 17.7, 6.5 and 3.1 % variation respectively for DFI, 2, 3, 4 and 5.

Sajina, Chakraborty, Jaiswar, Pazhayamadam, and Sudheesan (2011) analysed stock structure of *Megalaspis cordyla* (Linnaeus, 1758) based on principal component analysis and reported that the first three factors together explained 81.67 % of total morphometric variation. However in the present study seven principal components were extracted and it explains 93.15 % of variance among the six populations of *B. bendelisis* from two Himalayan Rivers. Hossain, Nahiduzzaman, Habiba, Mst, and Alam (2010) applied DFA and PCA on three populations of *Labeo calbasu* from the Jamuna, Halda and Hatchery Rivers and reported morphological discrimination among them, being attributed to the environmental factors and local migration of the fish. Khan, Miyan, and Khan (2012) also observed similar results in case of *Channa punctatus* from three rivers of India and concluded that environmental conditions have played a role in the movement, spatial distribution, and isolation of the fish stocks. Mir, Mir, and Chandra (2013) also pointed out that the closeness between the stocks may be due to their similar habitat attributes and environmental impacts. Some similar environmental parameters or the habitats could be the reason of closeness between the stocks. However the ecological factors were not studied in the present study. The variation in the populations of *B. bendelisis* of two Himalayan Rivers (Alaknanda and Chenab) is most probably because of the geographic isolation.

Two mitochondrial genes, 16S rRNA gene and the protein-coding COI genes highly conserved have been extensively studied in fish phylogenetics and have also been sequenced in various invertebrate and vertebrate taxa (Brown 1985; Bermingham & Lessios 1993; Santos, Schneider, & Sampaio, 2003; Munasinghe, Burr ridge, & Austin, 2004; Vinson, Grazielle, Schneider, & Sampaio, 2004; Ward, Zemlak, & Innes, 2005; Lakra, Goswami, & Gopalakrishnan, 2008). In the present study, the molecular divergence among the stocks of *Barilius bendelisis* was analysed with the help of COI gene which was mostly used in past studies for molecular characterization of fishes. All the studied sequences of COI in the present study were A+T rich, which was also reported earlier in fishes by Johns and Avise (1998). Sah et al. (2011) observed 8 different haplotypes among three populations of *B. Bendelisis* from Saryu, Kalsa and Kosi River, while in our study 11 haplotypes, (six haplotypes from the populations of Alaknanda River and five haplotypes for the population

of Chenab River) were found. Sah et al. (2011) also found 17 polymorphic sites, 14 parsimony informative sites in cytochrome b gene sequences in all three populations while in our case 7 polymorphic sites and six parsimony informative sites in the specimens from the Alaknanda River whereas four polymorphic sites and one parsimony informative site in the specimens from the Chenab River were found. Maximum likelihood tree shows 9 sequences of *B. bendelisis* from three different streams of Alaknanda forming one cluster and 9 other sequences of same species from three different streams of Chenab River forming other cluster. Thus both morphometric and molecular data support each other, in identification and phylogeny of *B. bendelisis* from two isolated rivers, Alaknanda and Chenab of India. The morphometric and molecular characterization of distinct populations of *B. bendelisis* is an important tool for conservation management and breeding programmes.

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## References

- Bermingham, E., & Lessios, H. A. (1993) Rate variation of protein and mtDNA evolution as revealed by sea urchins separated by the Isthmus of Panama. *Proceedings of the National Academy of Sciences of the United States of America*, 90, 2734-2738.
- Brown, W. M. (1985). The mitochondrial genome of animals. In R. J. McIntyre (Ed.), *Molecular Evolutionary Genetics* (pp. 95-130). New York: Plenum Press.
- Day, F. (1878). *Fishes of India* (Vol. 1 and Vol. 2). London: William Dawson and Sons Ltd.
- Elliott, N. G., Haskard, K., & Koslow, J. A. (1995). Morphometric analysis of orange roughy (*Hoplostethus atlanticus*) off the continental slope of southern Australia. *Journal of Fish Biology*, 46, 202-220.
- Hamilton, F. (1807). *An account of the fishes found in the river Ganges and its branches*. Edinburg: Archibald Constable and Company.
- Hammer, Ø., Harper, D. A. T., & Ryan, P. D. (2001). PAST: Paleontological statistics software package for education and data analysis. *Palaeontologia Electronica*, 4, 1-9.
- Hatcher, L. (1994). *A step by step approach to using the SAS system for factor analysis and structural modeling*. USA: SAS Institute.
- Hossain, M. A. R., Nahiduzzaman, M., Habiba, K. D. S., Mst, U., & Alam, M. S. (2010). Landmark-based morphometric and meristic variations of the endangered Carp, Kalibaus *Labeo calbasu*, from stocks of two isolated rivers, the Jamuna and Halda, and a hatchery. *Zoological Studies*, 49, 556-563.
- Hubert, N., Hanner, R., Holm, E., Mandrak, N. E., Taylor, E., & Burrige, M. (2008). Identifying Canadian freshwater fishes through DNA barcodes. *PLoS ONE*, 3(6), e2490. DOI: 10.1371/journal.pone.0002490

Humphries, J. M., Bookstein, F. L., Chernoff, B., Smith, G. R., Elder, R. L., & Poss, S. G. (1981). Multivariate discrimination by shape in relation to size. *Systematic Zoology*, 30, 291-308.

IUCN. (2012). *Red List of Threatened Species* (Version. 2012.1). *Journal of Fish Biology*, 74(2), 329-56.

Johns, G. C., & Avise, J. C. (1998). Tests for ancient species flocks based on molecular phylogenetic appraisals of *Sebastes* rockfishes and other marine fishes. *Evolution*, 52, 1135-1146.

Karakousis, Y., Triantaphyllidis, C., & Economidis, P. S. (1991). Morphological variability among seven populations of brown trout, *Salmon trutta L.*, in Greece. *Journal of Fish Biology*, 38, 807-817.

Khan, M. A., Miyan, K., & Khan, S. (2012). Morphometric variation of snakehead fish, *Channa punctatus*, populations from three rivers. *Journal of Applied Ichthyology*, 28, 154-155.

Kullander, S. O., Fang, F., Delling, B., & Ahlander, E. (1999). The fishes of the Kashmir Valley. In L. Nyman (Ed.), *River Jhelum, Kashmir Valley: Impact on the aquatic environment* (pp. 100-167). Sweden: SWEDMAR.

Lakra, W. S., Goswami, M., & Gopalakrishnan, A. (2008). Molecular identification and phylogenetic relationships of seven Indian Sciaenids (Pisces: Perciformes, Sciaenidae) based on 16S rRNA and cytochrome c oxidase subunit I mitochondrial genes. *Molecular Biology Reports*, 36, 831-839.

Menon, A. G. K. (1999). *Check list - Fresh water fishes of India*. Records of the Zoological Survey of India, Occasional Paper No. 175. India: ZSI.

Mir, F. A., Mir, J. I., & Chandra, S. (2013). Phenotypic variation in the Snowtrout *Schizothorax richardsonii* (Gray, 1832) (Actinopterygii: Cypriniformes: Cyprinidae) from the Indian Himalayas. *Contributions to Zoology*, 82(3), 115-122.

Mir, J. I., Sarkar, U. K., Dwivedi, A. K., Gusain, O. P., & Jena, J. K. (2013). Stock structure analysis of *Labeo rohita* (Hamilton, 1822) across the Ganga basin (India) using truss network system. *Journal of Applied Ichthyology*, 29(5), 1097-1103.

Mir, J. I., Saxena, N., Patiyal, R. S., & Sahoo, P. K. (2015). Phenotypic differentiation of *Barilius bendelisis* (Cypriniformes: Cyprinidae) in four rivers from Central Indian Himalaya. *Revista de Biología Tropical*, 63(1), 165-173.

Mirza, M. R. (1991). A contribution to the systematics of the Schizothoracine fishes (Pisces: Cyprinidae) with the description of three new tribes. *Pakistan Journal of Zoology*, 23, 339-341.

Munasinghe, D., Burridge, C. P., & Austin, C. M. (2004). Molecular phylogeny and zoogeography of the (Decapoda: Parastacidae) in Australia. *Biological Journal of the Linnaean Society*, 81(81), 553-563.

- Rahman, A. K. A. (1989). *Freshwater fishes of Bangladesh*. Dhaka: Zoological Society of Bangladesh.
- Rock, J., Costa, F. O., & Walker, D. D. (2008). DNA barcodes of fish of the Scotia Sea, Antarctica indicate priority groups for taxonomic and systematic focus. *Antarctic Science*, *20*, 253-262.
- Rohlf, F. J. (2006). *tpsDig2* (version 2.1). Department of Ecology and Evolution, State University of New York. Stony Brook, USA. Retrieved from <http://life.bio.sunysb.edu/morph/index.html>
- Rozas, J. & Rozas, R. (1999). DnaSP version 3: an integrated program for molecular population genetics and molecular evolution analysis. *Bioinformatics*, *15*, 174-175.
- Sah, S., Barat, A., Pande, V., Sati, J., & Goel, C. (2011). Population structure of Indian Hill Trout (*Barilius bendelisis*) inferred from variation in mitochondrial DNA sequences. *Advances in Biological Research*, *5*(2), 93-98.
- Sahoo, P. K., Saikia, S. K., & Das, D. N. (2009). Natural food resources and niche breadth of *Barilius bendelisis* (Hamilton) (Pisces, Cyprinidae) in river Dikrong, an upland riverine ecosystem in India. *Pan-American Journal of Aquatic Sciences*, *4*(1), 12-16.
- Sajina, A. M., Chakraborty, S. K., Jaiswar, A. K., Pazhayamadam, D. G., & Sudheesan, D. (2011). Stock structure analysis of *Megalaspis cordyla* (Linnaeus, 1758) along the Indian coast based on truss network analysis. *Fisheries Research*, *108*, 100-105.
- Sambrook, J., Fritsch, E. F., & Maniatis, T. (2001). *Molecular cloning: A laboratory manual* (2<sup>nd</sup> ed.). New York: Cold Spring Harbor Laboratory Press.
- Santos, S., Schneider, H., & Sampaio, I. (2003). Genetic differentiation of *Macrodon ancylodon* (Sciaenidae, Perciformes) populations in Atlantic coastal waters of South America as revealed by mtDNA analysis. *Genetics and Molecular Biology*, *26*(2), 151-161.
- Steinke, D., Zemlak, T. S., & Hebert, P. D. N. (2009). Barcoding Nemo: DNA based identifications for the ornamental fish trade, *PLoS One*, *4*(7), e6300.
- Strauss, R. E., & Bookstein, F. L. (1982). The truss: body form reconstruction in morphometrics. *Systematic Zoology*, *31*, 113-135.
- Swartza, E. R., Mwalea, M., & Hanner R. (2008). A role for barcoding in the study of African fish diversity and conservation. *South African Journal of Science*, *104*, 293-298.
- Talwar, P. K., & Jhingran, A. G. (1991). *Inland fishes of India and adjacent countries* (Vol. I & II). New Delhi: Oxford and IBH Publishing Company Pvt. Limited.
- Tamura, K., Stecher, G., Peterson, D., Filipowski, A., & Kumar, S. (2013). MEGA6: Molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, *30*, 2725-2729.

- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., & Higgins, D. G. (1997). The Clustal X windows interface: Flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Research*, *24*, 4876-4882.
- Turan, C. (1999). A note on the examination of morphometric differentiation among fish populations: the truss system. *Turkish Journal of Zoology*, *23*, 259-263.
- Victor, B. C., Hanner, R., Shivji, M., Hyde, J., & Caldow, C. (2009). Identification of the larval and juvenile stages of the *Cubera snapper*, *Lutjanus cyanopterus*, using DNA barcoding. *Zootaxa*, *2215*, 24-36.
- Vinson, C., Grazielle, G., Schneider, H., & Sampaio, I. (2004). Sciaenidae fish of the Caete river estuary, Northern Brazil: Mitochondrial DNA suggests explosive radiation for the Western Atlantic assemblage. *Genetics and Molecular Biology*, *27*(2), 174-180.
- Ward, R. D., Hanner, R., & Hebert, P. D. N. (2009). The campaign to DNA barcode all fishes, FISH-BOL. *Journal of Fish Biology*, *74*(2), 329-356.
- Ward, R. D., Zemlak, T. S., & Innes, B. H. (2005). DNA barcoding Australia's fish species. *Philosophical Transactions of the Royal Society B Biological Sciences*, *360*, 1847-1857.
- Wimberger, P. H. (1992). Plasticity of fish body shape the effects of diet, development, family and age in two species of *Geophagus* (Pisces, Cichlidae). *Revista de Biología Tropical*, *45*, 197-218.