

## Reintroducing plant coverage in a tropical wetland contaminated with oil and sulfate: rhizosphere effects on *Desulfovibrio* populations

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**Abstract:** The Mexican tropical wetland is a coastal system with capacity to support the contamination derived from the extractive industry and the transformation of crude oil, due to its high plant biodiversity and the presence of rhizospheric reducing sulphate bacteria from plants tolerant to crude oil. A field experiment was carried out for nine months to evaluate the adaptation of aquatic plants *Leersia hexandra* grass and *Eleocharis palustris* spikerush reintroduced in a wetland contaminated with 75 560 to 118 789 mg kg<sup>-1</sup> of weathered petroleum and also with sulfate, derived from oil and gas pipeline leaks, and gaseous emissions. The effect of the weathered oil and sulfate on the dry matter production and the population density of the bacterium *Desulfovibrio* spp, isolated from the rhizosphere and soil of both plants, were evaluated. The means of the variables had statistical differences ( $p < 0.05$ ). Weathered oil inhibited dry matter production of *L. hexandra* but not *E. palustris*; the effect of petroleum on *Desulfovibrio* density was very significant negative in the rhizosphere and in the soil of both plants. Sulfate reduced the dry matter of grass. The exposure of *Desulfovibrio* to sulfate significantly reduced its density in rhizosphere and soil ( $p < 0.01$ ). We recommend the use of *E. palustris* for the decontamination of flooded soils contaminated with weathered oil and sulfate. Rev. Biol. Trop. 66(2): 908-917. Epub 2018 June 01.

**Key words:** bacteria; Gleysol; *Eleocharis*; *Leersia*; weathered petroleum.

The extraction and transportation of crude oil is an environmental problem for the ecosystem (Muratova, Dmitrieva, Panchenko, & Turkovskaya, 2008) due to accidental spilling and subsequent permanence of oil in soils. In the Mexican humid tropics there are farms where oil has been spilled and then remains in the soil (Rivera-Cruz, Trujillo-Narcía, Trujillo-Rivera, Arias-Trinidad, & Mendoza-López, 2016). These farms make use of forage crops adapted to wetlands, where there is a predominance of stationary flooding, low permeability

of soils and shallow water cover (Comisión Nacional para el Conocimiento y Uso de la Biodiversidad [CONABIO], 2009). The initial spill of crude oil in the soil inhibits native life, but when it remains for a long period time it is colonized by organisms which are tolerant to hydrocarbons. In this regard Merkl, Schultze-Kraft & Infante (2005) and Basumatary, Bordoioi & Sarma (2012) demonstrated that tropical grass *Brachiaria brizantha*, and sedge *Cyperus brevifolius* are resistant to petroleum-polluted soils. The tropical grasses *Paspalum virgatum*

and *Echinochloa polystachya*, prevalent in contaminated wetlands, possessing rhizospheres colonized by beneficial bacteria (Rivera-Cruz, 2011). Plants and bacteria resistant to contamination are the basis of biological succession for the restoration of degraded lands in these ecosystems. Adaptation of grasses and sedges to stress conditions is due to two reasons. First, when the plant is exposed to a contaminant it activates its antioxidant enzyme system and molecular defense for its anatomical and physiological adaptation (Martí et al., 2009). Second, the fibrous structure, extensive root systems, root exudates and microbial density in the rhizosphere use available organic and inorganic molecules as sources of carbon and energy (Basumatary, Bordoloi, & Sarma, 2012; Rivera-Cruz, Maldonado-Chávez, & Trujillo-Narcía, 2012). In barium sulfate mines and soils contaminated with oil, and also acidophilous soils which predominate in farms in the humid tropics, there are sulfate-reducing bacteria (SRB) (Fernández, Rojas, & Roldán, 2006; Babu, Subramanyam, Sreenivasulu, & Paramageetham, 2014). The SRB are a unique physiological group of heterotrophic prokaryotes because they have the capability of using sulfate as the final electron acceptor in respiration. SRB are anaerobes which are characterized by their ability to perform dissimilatory sulfate reduction with the simultaneous oxidation of the organic substrates (Hansen, 1994). The SRB use electrons as donors for the reduction of hydrogen sulfate, ethanol, acetate, lactate, propionate, pyruvate (Liamleam & Annachatre, 2007) and hydrocarbons (Madigan, Martinko, Bender, Buckley, & Sthal, 2015). An example of SRB is the subgroup *Desulfovibrio* which is heterotrophic and capable of using the  $\text{SO}_4^{2-}$  or sulfur oxyanions as electron acceptors in the process of sulfate reductive assimilation to  $\text{H}_2\text{S}$  (Hurst, 2002). The reduction to  $\text{H}_2\text{S}$  by the sulfate reductive assimilation pathway is a natural *in situ* process under anaerobic conditions and is based on the following two fundamental reasons. Firstly, SRB have the capacity to reduce sulfate to sulfide, which then reacts with certain metals to form insoluble

precipitates. Secondly, the system acidity is reduced by their own action of sulfate reduction and by the carbon metabolism of the bacteria (García, Moreno, Ballester, Blázquez, & González, 2001). *Desulfovibrio* can inhabit soils contaminated with petroleum oil (Fernández, Rojas, & Roldán, 2006) but it also colonizes the rhizosphere of hydrophilic plants. The rhizosphere is critical for *Desulfovibrio* populations because it is the space where the roots stabilize organic and inorganic molecules by a radical adhesion process and organic and inorganic exudates (Dzantor, 2007). These exudates provide sufficient carbon and energy to support a large number of microbes in the rhizosphere (Erickson, Davis, & Narayanan, 1995). This plant-induced enhancement of the microbial population is called the rhizosphere effect (Atlas & Bartha, 2002). The rhizosphere of the grasses *E. polystachya* and *C. brevifolius* have shown a potential to be colonized by microorganisms and to degrade petroleum hydrocarbons (Rivera-Cruz & Trujillo-Narcía, 2004; Basumatary, Bordoloi, & Sarma, 2012; Rivera-Cruz et al., 2016). *L. hexandra* and *E. palustris* are potentially interesting species for the reestablishment of plant cover in oil contaminated soils. As both are mostly vegetatively propagated species, the best method for its establishment in these environments is planting rhizomes. *L. hexandra* has a fibrous and stoloniferous root system and creeping growth. *E. palustris* has taproots with tiller growth. Both root systems can control the spread of petroleum hydrocarbons in soils. The overall aim of this research was to evaluate the suitability of *L. hexandra* and *E. palustris* for recovering plant cover in a tropical wetland contaminated with oil, as well as to assess the effect of introduced vegetation on *Desulfovibrio* populations.

## MATERIALS AND METHODS

**Study area and environmental conditions:** Comprises 26 254 m<sup>2</sup> at La Venta, Tabasco, located in Southeast Mexico (UTM 389515 and 1998987) at 3 m elevation in a coastal plain. The climate is tropical wet with summer

rains, the average annual rainfall is 2200 mm and the mean annual temperature is 26 °C (Instituto Nacional de Estadística y Geografía [INEGI], 2001). The soil is Gleysol and the vegetation is mostly hydrophilic, dominated by *Typha domingensis*, *E. palustris*, *Cyperus surinamensis*, *Cynodon plectostachyus*, *Echinochloa polystachya*, *L. hexandra*, and *Paspalum virgatum*. Oil infrastructure includes nine pipelines from 6 to 20 inches in diameter, carrying crude oil, gas and ethane, is also a dam with residues of drilling an oil well. The spills of crude oil arrived at the farm via ruptured pipelines connecting the wells with storage batteries and La Venta Gas Processing Complex.

**Oil spill and study areas:** The oil spill occurred in 2006 and differently affected the farm depending on the distance to the contamination focus. Drilling was done with auger closed up to one meter deep in the soil. Four areas with different degrees of environmental impact were identified due to the chronic oil spill. Each area (A) consisted of a surface of 30 m<sup>2</sup> (5x6 m). The area was cleared of native vegetation, and the soil was removed, crushed and homogenized. Soil was randomly collected from 10 m<sup>2</sup> in each area leaving an effective border of 1.0 m width in order to evaluate the Total Petroleum Hydrocarbons in soil (TPHs) contents. The oil contents were determined using the modified soxhlet extraction method of EPA 3540B (United States Environmental Protection Agency [USEPA], 1994). The samples were acidified with HCl to pH 2 and then dried with MgSO<sub>4</sub>. Soxhlet extraction was undertaken for 8 h using dichloromethane of 99.5 % purity (Merck® analysis). The solvent was evaporated and the dry weight of the extract was quantified in mg kg<sup>-1</sup> (gravimetry).

**Experimental design and setup:** The experiment was conducted as a 4x2 two factor factorial with four replicates: the first factor being TPHs content in soil (four levels: 912, 75560, 91695 and 118879 mg kg<sup>-1</sup>), and the second one the plant type (two levels). In September 2014, 10 cm long fragments of

stems with mature and differentiated buds of *L. hexandra* grass and *E. palustris* rhizomes collected in adjacent uncontaminated areas (300 of each species, equally distributed) were planted. Each area of 30 m<sup>2</sup> was delimited with square wire-mesh fencing to prevent any grazing animals from invading the test sites. Neither fertilization nor watering was implemented during the growing period.

**Foliar, soil and rhizosphere sampling:**

Nine months after the establishment of the experiment, four *one square meter replicates were randomly established in each zone*. Foliar biomass of both plant species was collected and dry matter production (DMP) was determined. Also, for each plant species, samples of bulk and rhizosphere soil (soil firmly adhered to roots) from individual plants were collected for assessments of assimilable sulfate (SO<sub>4</sub><sup>-2</sup>) and bacteria populations of subgroup *Desulfovibrio*.

**Dry matter production:** The aerial biomass (stems and leaves) of *L. hexandra* and *E. palustris* were washed with distilled water and were oven dried at 65 °C until constant weight determination.

**Sulfate analysis:** Bulk and rhizosphere soil samples from *L. hexandra* and *E. palustris* were dried, ground and sieved, and the assimilable SO<sub>4</sub><sup>-2</sup> was determined with a buffer solution (K<sub>2</sub>SO<sub>4</sub>, 2N glacial acetic acid and distilled H<sub>2</sub>O) and a standard solution [Ca(H<sub>2</sub>PO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O, 2N glacial acetic acid and distilled H<sub>2</sub>O]. Sulfate quantification was achieved by the turbidimetric method in a spectrophotometer at 670 nm (Etchevers, 1992).

**Desulfovibrio population density:** Bulk and rhizosphere soil of *L. hexandra* and *E. palustris* were stored at 4 °C for one week prior to analysis of the population of SRB (*Desulfovibrio*) which was evaluated in a specific culture medium (ML) under anaerobic conditions; ML medium was composed of the following (g/L): Bacto agar™ 10, Bacto tryptone agar™ (tryptone 8, peptone 2, dextrose

15, agar 15 and bromocresol purple 0.04), 10 Na<sub>2</sub>SO<sub>3</sub>, FeSO<sub>4</sub>(NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>·6H<sub>2</sub>O at 5 %, 10 Buffer solution (g/L): MgCl 0.811 and KH<sub>2</sub>PO<sub>2</sub> 0.34. The medium pH was adjusted (7.6 ± 0.1) previously and transferred into tubes with rubber lids perforated by metallic needles and autoclaved for 15 min at 120 °C (ASTM, 2009). The system was depressurized for oxygen removal through the needles, which were rapidly removed after autoclaving. The samples were inoculated using 1.0 mL each injected into 9 mL of the ML medium, through the metallic needles. After incubating in an anaerobic incubator for 48 h at 37 °C, positive tubes for SRB showed black iron sulfide deposition (Fernández et al., 2006). To calculate the CFU/g we used the most probable number with decimal dilutions and five tubes per dilution (Cochran, 1950).

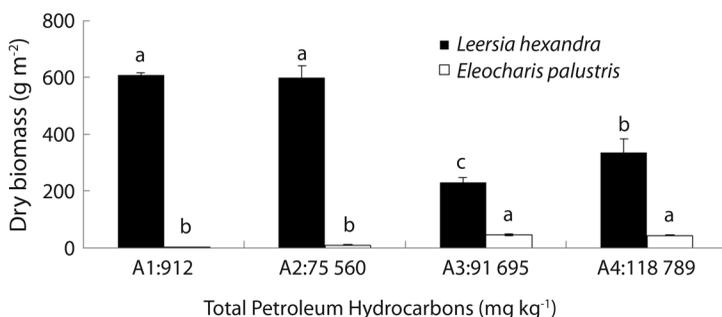
**Statistical analysis:** The analysis of variance (ANOVA) of the factors (TPHs content and plant type) was calculated with respect to the parameters of the conducted study using the statistical program SAS version 8.01 (SAS, 2000). DMP, SO<sub>4</sub><sup>-2</sup> and populations of *Desulfovibrio* in bulk and rhizosphere soil of *L. hexandra* and *E. palustris* were subjected to one way ANOVA in order to test for significant differences between treatments (p < 0.05). Moreover, the pairwise correlation between DMP, TPHs, sulfate and *Desulfovibrio* in soil and rhizosphere in the contaminated treatments of both vegetal species was evaluated with Pearson's

correlation-coefficient, using linear regression, for normally distributed variables with a statistical significance of p < 0.05.

## RESULTS

**Total Petroleum Hydrocarbons in soil (TPHs):** The initial contents of TPHs (mg kg<sup>-1</sup>) in each area were A1: 912 ± 230, A2: 75 560 ± 13 387, A3: 91 695 ± 19 467, and A4: 118 789 ± 18 623. The soils are strongly acidic (pH H<sub>2</sub>O from 4 to 4.6), clay loam texture, with 5.9, 9.5, 11.4 and 13.4 % organic carbon, respectively.

**Dry matter production:** Contents of TPHs, plant type and their interaction were statistically significant. Dry matter production (DMP) of *L. hexandra* was significantly higher than of *E. palustris* independently of the TPHs content. Higher values for *L. hexandra* were reached with lower TPHs contents (A1 and A2), the DMP for *E. palustris* significantly increased with the higher TPHs contents (Fig. 1). For *L. hexandra*, a significant negative correlation was found between dry matter and both TPHs and sulfate contents in rhizosphere soil. By the contrary, DMP was significantly and positively correlated with *Desulfovibrio* populations in bulk soil. The dry matter of *E. palustris* was positive and significantly correlated with TPHs and sulfate contents in bulk soil, and negatively correlated with *Desulfovibrio* populations both in bulk and rhizosphere soil (Table 1).



**Fig. 1.** Dry matter of *Leersia hexandra* and *Eleocharis palustris* after nine months growing in soil with different TPHs content. Each column represents the mean of four replications; bars indicate standard deviation. Means for each plant type followed by different letters are statistically significant at p < 0.05 as determined by Tukey's test.

TABLE 1

Pairwise correlation coefficients and test for zero pairwise Pearson correlation of the corresponding variables at the ninth month of the experiment for *Leersia hexandra* and *Eleocharis palustris* in soils with TPHs contents

|  | TPHs    | SO <sub>4</sub> <sup>-2</sup> -bulk soil | SO <sub>4</sub> <sup>-2</sup> -rhizosphere | Dv - bulk soil | Dv - rhizosphere |
|--|---------|--|--|----------------|------------------|
| <i>Leersia hexandra</i>                                |         |  |  |                |                  |
| <sup>1</sup> DMP                                       | -0.690* | -0.444                                   | -0.623*                                    | 0.598*         | 0.244            |
| <sup>2</sup> TPHs                                      |         | 0.777**                                  | 0.984**                                    | -0.987**       | -0.843**         |
| <sup>3</sup> SO <sub>4</sub> <sup>-2</sup> - bulk soil |         |  | 0.835**                                    | -0.740**       | -0.858**         |
| SO <sub>4</sub> <sup>-2</sup> - rhizosphere            |         |  |  | -0.971**       | -0.887**         |
| <sup>4</sup> Dv - bulk soil                            |         |  |  |                | 0.881**          |
| <i>Eleocharis palustris</i>                            |         |  |  |                |                  |
| DMP  | 0.821** | 0.694**                                  | 0.466                                      | -0.727**       | -0.689**         |
| TPHs   |         | 0.908**                                  | 0.698**                                    | -0.969**       | -0.948**         |
| SO <sub>4</sub> <sup>-2</sup> - bulk soil              |         |  | -0.824**                                   | -0.790**       | -0.489           |
| SO <sub>4</sub> <sup>-2</sup> - rhizosphere            |         |  |  | -0.723**       | -0.732**         |
| Dv - bulk soil   |         |  |  |                | 0.995**          |

<sup>1</sup>Dry matter production, <sup>2</sup>Total Petroleum hydrocarbons, <sup>3</sup>Sulfate, <sup>4</sup>*Desulfovibrio*, \*Significant at p< 0.05 and \*\*Significant at p< 0.01.

**Sulfate contents:** Contents of SO<sub>4</sub><sup>-2</sup> in bulk and rhizosphere soil of *L. hexandra* and *E. palustris* were significantly different among contamination levels (Fig. 2). In general, sulfate contents in rhizosphere soil was significantly higher than in bulk soil with the exception of *L. hexandra* growing in the less contaminated area. There were no significant differences between plant types (Table 1). For *L. hexandra*,

sulfate contents in bulk and rhizosphere soil were significantly and positively correlated between them and with TPHs. Also, sulfate contents were negative and significantly correlated with *Desulfovibrio* populations in bulk and rhizosphere soil (Table 1). For *E. palustris*, sulfates in bulk and rhizosphere soil were negative and significantly correlated between them, and with *Desulfovibrio* populations in bulk

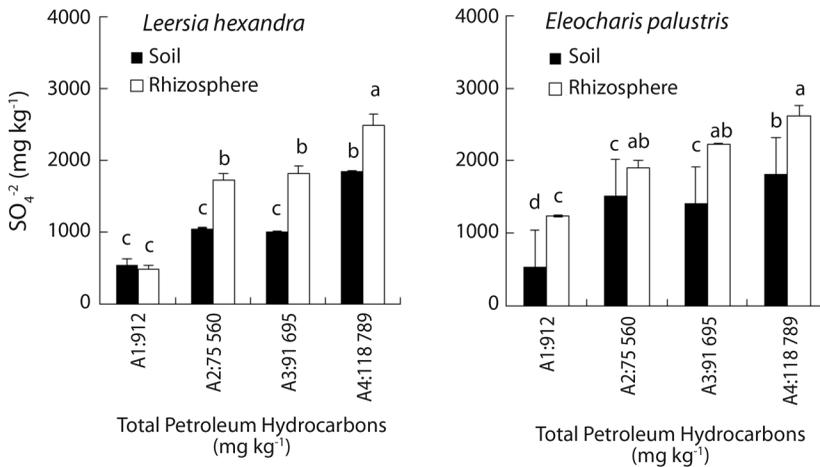
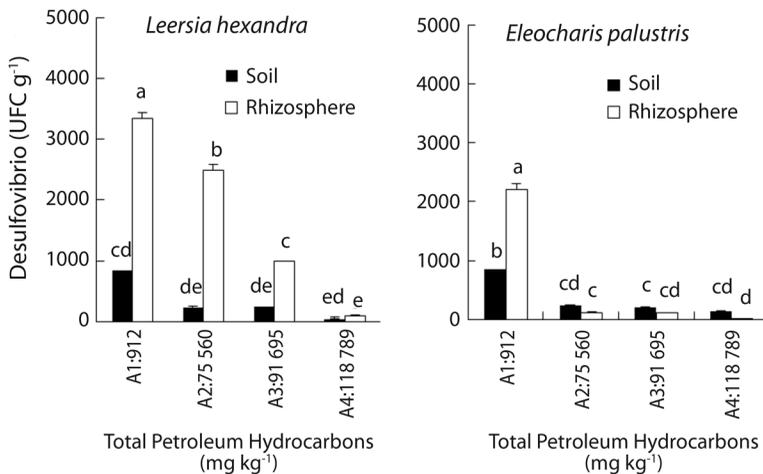


Fig. 2. Sulfate in bulk and rhizosphere soil of *Leersia hexandra* and *Eleocharis palustris* after nine months growing in soil with different TPHs contents. Each column represents the mean of four replications; bars indicate standard deviation. Means not sharing letters are statistically different at p< 0.05 as determined by Tukey's test.





**Fig. 3.** Desulfovibrio in bulk and rhizosphere soil of two plant species, after nine months growing in soil with different TPHs content. Each column represents the mean of four replications; bars indicate standard deviation. Means not sharing letters are statistically different at  $p < 0.05$  as determined by Tukey's test.

soil. Sulfates and *Desulfovibrio* populations, both in rhizosphere soil, were also negatively and significantly correlated (Table 1).

**Desulfovibrio population density:** *Desulfovibrio* populations in *L. hexandra* were always significantly higher in rhizosphere than in bulk soil. For this plant higher *Desulfovibrio* amounts were obtained in the less contaminated soil, diminishing progressively with higher oil contents. In *E. palustris* the rhizosphere effect on *Desulfovibrio* populations was only significant with the lower oil contents, where rhizosphere populations were higher than in bulk soil. With higher oil contents *Desulfovibrio* populations diminished considerably, with no differences between bulk and rhizosphere soil (Fig. 3). *Desulfovibrio* populations in bulk and rhizosphere soil of both plants tested were negatively and significantly correlated with TPHs, and positively and significantly correlated between them (Table 1).

## DISCUSSION

Different studies have shown the inhibitory effect of oil on plant growth (Merkl, Schultze-Kraft, & Infante, 2004; Rivera-Cruz,

Trujillo-Narcía, Miranda, & Maldonado, 2005; Hou, Liu, Zhang, Hu, & Cao, 2016), although in some cases it is different. In this research, the DMP production of *L. hexandra* significantly decreased with higher TPHs contents. Similar inhibitory results were found by Rivera-Cruz & Trujillo-Narcía (2004) in the grasses *E. polystachya* and *B. mutica*, in a Gleysol with TPHs of 79 457 mg kg<sup>-1</sup> who indicated that the DMP of both species decreased by 32 and 44.1 %, respectively. They attributed such behavior to lower height, thin stems achieved by the grass and some runners falling on the soil and not being rooted enough to produce tillers, thus leaving gaps. Likewise found that *Eleusine indica* biomass decreased by 46.8 % in soil with 5 % oil (Merkl et al., 2005).

One explanation they describe is that the life cycle of this species is delayed in the presence of TPHs, because they affect the amount of plant growth regulators, hence suggesting that patterns of plant growth are affected by the oil contained in the soil. Classic studies (Plice, 1949; Baker, 1971) pointed out that some plants can increase their growth in soils with oil. In our research, the biomass production of *E. palustris* spikerush showed a direct proportional response to the increase of the oil in the soil, whereas the ratio was inversely related

to the production of *L. hexandra* biomass. Oil inhibited the growth of this grass. The results of DMP in *E. palustris* shows a positive trend with the increase of the amount of oil in the soil. It resembles the effect of hormesis that occur in plants adapted to stressors by exposure to a pollutant during long periods of the life cycle (Calabrese, Baldwin, & Holland, 1999). According to Vaziri et al. (2013) the higher production of dry vegetable matter can be effective because of the greater exploration of the radical system and also the radical exudates that improve the catabolic activity of the rhizospheric microorganisms. One type of adaptation of plants growing on oil-contaminated soils is the increase in quantity and thickness of the root, it covers a larger area of exploration. *E. palustris* has morphological and physiological characteristics that stimulate the adaptation and growth of the plant to unfavorable conditions (Hauser, 2006; Tilley & John, 2012). It is a pioneer species that covers the clay soil very quickly as the water level decreases, and also under flood conditions, it is dominant in all serous stages. It is early seral because of the rapid rhizomatous growth in warm environments, both in soil with water table at 30 cm below the surface, also in flooded terrains. According Tilley & John (2012) the elongation of the rhizome follows an almost linear trajectory to escape the competition of the same species. The same authors also mention that *E. palustris* has a dense root mass that extends more than 40 cm in the depth of the soil. It thrives in disturbed places. Species used for restoration of wetlands. It can fix atmospheric nitrogen and make it available to other plants in the wetland community (Tilley & John, 2012). This set of vegetal attributes may be related to the increase of the vegetal biomass in soils contaminated with greater amounts of petroleum. Only when TPHs contents partially inhibited the growth of *L. hexandra*, it was that *E. palustris* increased.

There was a significant correlation between TPHs and sulfate contents both in bulk and rhizospheric soil. The presence of high amounts of sulfur in soils adjacent to

the study area was reported by (Fernández et al., 2006), and the increases of sulfate in the study area may be due to the high content of sulfuric-containing organic molecules (sulfide, disulfide, mercaptan, thiophene, benzothio- phene and dibenzotione derivatives of resins and asphaltenes) (Waiquer, 2004) contained in the oil spill, which could have been released through mineralization processes. The accumulation of  $\text{SO}_4^{2-}$  was stimulated in both rhizospheres evaluated in this research and this could be explained by the presence of the roots, introducing oxygen and inducing the establishment of chemoautotrophic bacteria which oxidize hydrogen sulfide ( $\text{H}_2\text{S}$ ), elemental sulfur ( $\text{S}_0$ ) and thiosulfate ( $\text{S}_2\text{O}_3^{2-}$ ) in the soil (Lynch & Hobbie, 1988). Also, could partially be explained by the mineralization of soil organic carbon and lysis of microorganisms. The effect of the TPHs content, plant type and interactions were significant for *Desulfovibrio* in rhizosphere soil, where populations of these bacteria were always higher in *L. hexandra* than in *E. palustris*. The type of plant had no effect on the bulk soil.

According to Alexander (1994) sulfate reducing bacteria in acidophilus anoxic environments saturated with S, occur naturally in amounts between  $10^2$  to  $10^4$  CFU per gram of soil. In our research the population subgroup *Desulfovibrio* fitted this range in bulk and rhizospheric soil of *L. hexandra*, while these bacteria populations in *E. palustris* were considerably lower (Fig. 3). In rhizosphere soil of *L. hexandra*, there was a significant increase in *Desulfovibrio* populations with regard to bulk soil, this rhizosphere effect has been reported to be favored by pastures (like *L. hexandra*) over other plant species, as noted by Badalucco & Kuikman (2001) in *Holcus lanatu*, *Anthoxanthum odoratum*, *Festuca rubra*, and *Festuca ovina*. Sacaca, Iñiguez & Roulet, (2009) reported an increase in populations and selectivity of sulfate reducing bacteria belonging to the subgroup *Desulfobacter* in roots of aquatic plants. In our study the rhizosphere of *L. hexandra* induced proliferation of *Desulfovibrio* to a greater extent than *E. palustris*.

This response could be attributable to the type of fibrous root system, extensively characterizing *L. hexandra*. This root architecture has been linked to an increase in the availability of  $\text{SO}_4^{2-}$  in different pastures growing in anoxic environments (Aprill & Sims, 1990), which is also a characteristic of our study area. The presence of *Desulfovibrio* in these environments is linked to its intervention in processes of nutritional sulfate assimilation. Madigan et al. (2015) noticed that *Desulfovibrio sulfofidismutans* promotes the reduction of sulfur compounds using an intermediate oxidation state, wherein one compound is oxidized and another is reduced, thus reducing thiosulfate ( $\text{S}_2\text{O}_3^{2-}$ ), sulfite ( $\text{SO}_3^{2-}$ ) and sulfur ( $\text{S}_0$ ).

Hence, an atom of sulfur thiosulfate is oxidized ( $\text{SO}_4^{2-}$ ) and another is reduced (forming  $\text{H}_2\text{S}$ ). The small population densities of *Desulfovibrio*, in the soil and in the rhizosphere of *L. hexandra* and *E. palustris*, are of few importance in the elimination of petroleum in the soil, this explains why this bacterium uses a limited number of sources of C (Parés & Juárez, 1997). This low spectrum of usable substrates is because the sulfate oxidation has low energy yield, therefore the energy that can be obtained from the oxidation of the substrate is small (Madigan et al., 2015; Parés & Juárez, 1997). It is known that the rate of biodegradation of organic compounds under anaerobic conditions is lower than aerobic conditions caused by the lower amount of energy sources, consequently decreasing the multiplication of new generations of bacteria (Madigan et al., 2015; Parés & Juárez, 1997; Santos, Carmo, Pases, Rosado, & Peixoto, 2011), even the presence of sulphate in the soil and rhizosphere inhibited *Desulfovibrio* density in both plants (Table 1). The greater permanence of the oil in flooded soils may be precisely due to the decrease of the microbial activity, so that the persistence of the contaminant can last for decades. The reintroduction of the vegetation cover in a tropical wetland contaminated with petroleum and sulfate evidenced different effects in the growth and development of *L. hexandra* grass and in *E. palustris* spikerush. *Desulfovibrio* population's

densities were higher in rhizospheres of both plant species but decreased in soils contaminated with petroleum and sulfate. *E. palustris* plant showed adaptation during the nine months of exposure to oil and sulfate, vegetative growth was not inhibited. We recommend the use of *E. palustris* for the decontamination of flooded soils contaminated with weathered oil and sulfate.

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## RESUMEN

**Reintroducción de cobertura vegetal en un humedal tropical contaminado con petróleo y sulfato: efecto de la rizosfera sobre poblaciones de *Desulfovibrio*.** El humedal tropical mexicano es un sistema costero con capacidad para resistir la contaminación derivada de la industria extractiva y la transformación del petróleo crudo, debido a su alta biodiversidad de plantas y la presencia de bacterias rizosféricas reductoras de sulfato de plantas tolerantes al petróleo crudo. Se realizó un experimento en campo durante nueve meses para evaluar la adaptación de las plantas acuáticas *Leersia hexandra* y *Eleocharis palustris* reintroducidas en un humedal contaminado con 75 560 a 118 789  $\text{mg kg}^{-1}$  de petróleo intemperizado y también con sulfato, derivados de fugas de oleoductos y de emisiones gaseosas crónicas. Se evaluó el efecto del petróleo intemperizado y del sulfato en la producción de materia seca vegetal y en la densidad poblacional de la bacteria *Desulfovibrio* spp, aislada de la rizosfera y del rizoplano de ambas especies vegetales. Las medias de las variables tuvieron diferencias estadísticas ( $p < 0.05$ ). El petróleo intemperizado inhibió la producción de materia seca de *L. hexandra* pero no de *E. palustris*; el efecto del petróleo en la densidad de *Desulfovibrio* fue negativo muy significativo en la rizosfera y en el rizoplano de ambas plantas. El sulfato redujo la materia seca de *L. hexandra*. La exposición de *Desulfovibrio* a sulfato redujo muy significativamente ( $p < 0.01$ ) su densidad en rizosfera y en rizoplano. Recomendamos el uso de *E. palustris* para la descontaminación de suelos inundables afectados con petróleo intemperizado y con sulfato.

**Palabras clave:** bacteria; Gleysol; *Eleocharis*; *Leersia*; petróleo intemperizado.

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